

Increased mRNA Expression of α_{2A} -Adrenoceptors, Serotonin Receptors and μ -Opioid Receptors in the Brains of Suicide Victims

Pablo V Escribá*, Andrés Ozaita^{1,2} and Jesús A García-Sevilla¹

¹Laboratory of Neuropharmacology, Associated Unit of the Cajal Institute (CSIC), Department of Biology, Institut Universitari d'Investigació en Ciències de la Salut (IUNICS), University of the Balearic Islands, Palma de Mallorca, Spain

The development of new therapies for the treatment of psychiatric disorders requires an in-depth knowledge of the molecular bases underlying these pathologies, which remain largely unknown. Alterations in adrenoceptors, serotonin receptors, and other G protein-coupled receptors (GPCRs) have been associated with suicide and depression. However, to date, there is little information about mRNA expression of the GPCRs in the frontal cortex of suicide victims. Our goal was to study the expression in the brain of these receptors. For this purpose, we measured mRNA levels by RT-PCR. We found that the expressions of α_{2A} -adrenoceptors, 5-HT_{1A}, 5-HT_{2A} serotonin receptors, and μ -opioid receptors were elevated in the post-mortem brains of these suicide victims with respect to matched controls. Moreover, in the case of α_{2A} -adrenoceptors (the only for which these data were available), a significant correlation was observed between the level of mRNA and protein quantified in the brain of the same subjects, indicating that protein synthesis of this receptor was not influenced by post-translational regulatory mechanisms. In addition, the degree of adrenoceptor and 5-HT receptor expressions appeared to be correlated in the brains of suicide victims and control subjects. Alterations in the expression of adrenoceptors, serotonin, and opioid receptors indicate that these signaling proteins might be related to the etiopathology of suicidal and depressive behaviors. Alternatively, such changes may represent adaptive mechanisms to compensate for other as yet unknown alterations. The results also suggest that these receptors could share common regulatory mechanisms.

Neuropsychopharmacology (2004) 29, 1512-1521, advance online publication, 16 June 2004; doi:10.1038/sj.npp.1300459

Keywords: human brain; GPCR; neurotransmitter receptor; suicide

INTRODUCTION

Molecular and functional alterations of neurotransmitter receptors and their norepinephrine and/or serotonin (5-HT) signaling pathways are believed to play a direct role in the pathogenesis of suicide and depression (Mann, 1999; Ressler and Nemeroff, 1999). On the other hand, a wide variety of G protein-coupled receptors (GPCRs; eg α_2 -adrenoceptors, 5-HT receptors, μ -opioid receptors, GABA_B receptors, cholinergic muscarinic M_2/M_4 receptors, etc.) serve as auto- or heteroreceptors, and control neurotrans-

*Correspondence: PV Escribá, Laboratory of Molecular and Cellular Biomedicine, Associated Unit of the Instituto de la Grasa (CSIC), Department of Biology, IUNICS, University of the Balearic Islands, 07122 Palma de Mallorca, Spain, Tel: +34 971 173433, Fax: +34 971 173184, E-mail: pablo.escriba@uib.es

²Current address: Laboratori de Neurofarmacologia, Facultat de Ciencies de la Salut i de la Vida, Universitat Pompeu Fabra; 08003, Barcelona, Spain.

Received 9 April 2003; revised 3 August 2003; accepted 3 September 2003

Online publication: 15 March 2004 at http://www.acnp.org/citations/Npp03150403402/default.pdf $\,$

mitter release (Schlicker and Göthert, 1998; Starke, 1981). There is evidence that the density of some of these GPCR proteins, such as α_2 -adrenoceptors and 5-HT receptors, is altered in the brains of suicide victims and in the platelets of depressed patients (Callado et al, 1998; De Paermentier et al, 1997; García-Sevilla et al, 1986, 1999; González et al, 1994; Meana et al, 1992; Meana and García-Sevilla, 1987; Mendelson, 2000; Ordway et al, 1994; Pandey et al, 1995; Sargent et al, 2000; Stockmeier et al, 1998). These alterations have been correlated with impaired neurotransmitter release that results from α_2 -adrenoceptor supersensitivity and/or serotonin receptor dysfunction. Thus, stimulation of presynaptic α_2 - and 5-HT-autoreceptors induce a decrease in the synthesis and release of norepinephrine and 5-HT (Esteban et al, 1999). Further evidence of the involvement of α_2 -adrenoceptors and 5-HT receptors in the pathophysiology of suicide and depression is that chronic antidepressant administration reduces the density and activity of these receptors in rat brain (Esteban et al, 1999; Invernizzi et al, 2001; Smith et al, 1981; Sibug et al, 1998; Subhash and Jagadeesh, 1997), and in the platelets of depressed subjects (García-Sevilla et al, 1986; Stahl, 1994; Sargent et al, 1997). Based on these observations, it has been suggested that patients with mood disorders might suffer an underlying disturbance of noradrenergic and/or serotonergic function. In this context, it has been shown that the main metabolite in the production of 5-HT (5-hydroxyindolacetic acid) is slightly decreased in the cerebrospinal fluid of suicide victims (Nordstrom et al, 1994; Roy et al, 1986). Finally, μ-opioid receptors have been implicated in the behavioral alterations associated with stress. Indeed, its mRNA expression is augmented in the brain of rats submitted to stress (Yamamoto et al, 2003) and its protein levels were increased in the post-mortem brains of suicides (Gross-Isseroff et al, 1990; Gabilondo et al, 1995).

Most studies of the biochemical alterations of neurotransmitter receptors during major depression have relied upon assays of radioligand binding. However, the functional state of GPCR proteins (which determines radioligand binding) is modulated by several factors, such as coupling to G proteins, desensitization by protein phosphorylation (mediated by kinases such as GRK, PKC, and PKA), sequestration, and internalization (Chuang et al, 1996). Therefore, the results of radioligand binding studies in the brain of suicide victims will not necessarily reveal alterations in these and other molecular events. This study was designed to quantify the mRNA levels for α_{2A} -adrenoceptors, 5-HT_{1A}, 5-HT_{2A}, and μ -opioid receptors in the prefrontal cortex of suicide victims since mRNA levels are not modulated by the above phenomena. For this purpose, reverse transcription followed by polymerase chain reaction (RT-PCR) was used under conditions that permitted quantitative assessment of mRNA cellular concentrations in post-mortem human samples. This technique has the advantage over other approaches to evaluate gene expression in that it is more sensitive, quantitative, and overcomes moderate RNA degradation. As a result, we show that there is a significant increase in the expression of the GPCRs transcripts analyzed in post-mortem brains of suicide victims. As these receptors share signaling mechanisms and common regulatory transcription sequences, we also studied the association between the levels of mRNA expression of the same receptors in the same brains. The results suggest that these receptors could be involved in the pathophysiology of depressive/suicidal behavior and their transcription regulation might be controlled by similar (adrenoceptors and 5-HT receptors) or divergent (opioid receptors) mechanisms.

METHODS

Subject Selection and Brains Samples

This study was approved by the Research Ethics Committee at the University of the Balearic Islands. Human brains were obtained at autopsy from the Institute of Anatomical Forensics (Palma de Mallorca, Spain). Specimens of prefrontal cortex (Brodmann's area 9, middle frontal gyrus) were dissected on a glass surface cooled with ice and care was used to include only gray matter (no more than a 1-mm white-matter border was included) in cortical samples. The suicide group consisted of 12 suicide victims (eight men and four females; mean age \pm SEM: 54.1 ± 4.4 years; postmortem delay, 26.2 ± 2.6 h; Table 1). The cause of death of the suicide victims, as determined by the medical examiner, was trauma caused by jumping from a height (n = 5), hanging (n=3), self-inflected gunshot wound (n=1), poisoning (n=1, carbon monoxide), drug overdose (n=1, fluoxetine), and electrocution (n=1). Brain pH was not measured, and no neuropathological exams were performed. In this group of suicide victims, a retrospective review of the antemortem medical histories was performed to determine whether the victims had been clinically diagnosed or were under drug treatment for psychiatric disorders. This analysis was conducted after the completion of the assays and was performed without access to the biochemical findings. Most suicide victims used in this study had a pre-mortem diagnosis of major depression (9/ 12) (Diagnostic and Statistical Manual of Mental Disorders, third edition, revised [DSM-III-R] criteria). A pre-mortem

Table I Characteristics of the Individual Subjects Included in This Study

ID	Age (year)	Sex	PMD ^a (h)	Psychiatric diagnosis	Cause of death
Suicides					
SI	30	М	20	MD^{b}	Gunshot wound
52	69	М	28	MD	Jumping from a height
S3	46	F	22	MD	Poisoning
S4	25	М	24	MD	Jumping from a height
S5	64	F	24	MD	Drug overdose
S6	70	F	48	MD	Jumping from a height
S7	51	М	24	MD	Hanging
S8	47	М	11	MD	Electrocution
S9	62	F	26	MD	Jumping from a height
\$10	69	М	28	Unknown	Hanging
SII	67	М	36	Unknown	Hanging
S12	50	М	24	Unknown	Jumping from a height
					, 1 0 0
Mean	54.1	4F/8M	26.2		
SEM	4.4		2.6		
Controls					
CI	30	Μ	30	None	Motor-vehicle accident
C2	70	Μ	24	None	Myocardial infarction
C3	42	F	44	None	Drowning
C4	26	Μ	8	None	Motor-vehicle accident
C5	60	F	24	None	Motor-vehicle accident
C6	75	F	36	None	Motor-vehicle accident
C7	50	Μ	24	None	Drowning
C8	47	Μ	24	None	Cirrhosis
C9	50	F	24	None	Motor-vehicle accident
CI0	65	Μ	21	None	Drowning
CII	65	Μ	36	None	Myocardial infarction
Mean	52.7	4F/7M	26.8		
SEM	4.8		2.8		

^aPMD: post-mortem delay.

^bMD: major depression.



diagnosis was not available in three subjects. In suicide victims (n=6), a toxicological screening (quantitative assays for antidepressant and other psychotropic drugs, and ethanol) was performed on blood, urine, liver, and kidney samples at the Toxicology Unit, Institute of Forensic Medicine, using standard procedures. In this group of suicide victims, ethanol was detected in five suicide subjects and antidepressant drugs (clomipramine, fluoxetine, and mianserin) were detected in four depressed suicides. However, neither the above drugs nor electrocution or carbon monoxide influenced the transcription rate of adrenoceptors, serotonin receptors, or μ -opioid receptors, except alcohol and fluoxetine that induced reductions in 5-HT_{1B} mRNA (Neumaier et al, 1996; Vazquez et al, 2002). In six suicides no toxicological screening data were available. The control group for this series of suicide victims consisted of similar brain specimens from 11 subjects (seven men and four females; mean \pm SEM: age, 52.7 \pm 4.8 years; postmortem delay, 26.8 + 2.8 h; Table 1) who died of different accidental or natural causes. None of the control subjects had documented history or family reports of psychiatric disorders or drug abuse. Suicide victims and control subjects were matched according to the following criteria: (1) gender, (2) age, and (3) post-mortem delay. The mean difference in age between suicide and control for individual matches (absolute values) was 2.4 ± 0.8 years, whereas the mean age difference between the groups was 1.4 ± 4.4 years. The mean difference in post-mortem delay between suicide and control subjects for individual matches was $7.6 \pm 1.9 \,\mathrm{h}$ whereas the mean difference between groups was 0.6 ± 2.5 h. In all, 10 control subjects and suicide victims were paired directly, and one control subject was paired with two suicides.

RNA Isolation from Human Brain Tissue

RNA was isolated from human brain samples by the acid guanidinium thiocyanate/phenol/chloroform extraction method (Chomczynski and Sacchi, 1987). Briefly, approximately 500 mg of human brain cortex was homogenized with 10 volumes of TRIzol (Life Technologies, Grand Island, NY, USA) in a blade-type homogenizer (Ultraturrax, Janke

& Kunkel, GMBH & Co KG, Staufen, Germany). After incubating for 5-min at room temperature, 1 ml of chloroform was added to the homogenate, which was agitated vigorously for 15 s and then incubated for 3 min at room temperature. This mixture was centrifuged for 10 min at $12\,000\,g$ and 4° C, and the upper (aqueous) phase was recovered and mixed with 2.5 ml of isopropyl alcohol, before incubating for 10 min at room temperature. This mixture was centrifuged for 10 min at 12 000 g and 4°C, the supernatant was removed, and the RNA pellet was washed with 5 ml of 75% ethanol before centrifuging for 5 min at 7500 g and 4°C. The DNA residue was then digested with RNase-free DNase (bovine pancreas, Boehringer) following the directions provided by the manufacturer. After phenol extraction and ethanol precipitation, the final pellet was resuspended in 100 µl of RNase-free water and stored at -80° C. The RNA concentration in these samples was usually between 1.5 and 3 µg/µl (as determined by absorption at 260 nm). Finally, the RNA samples were analyzed by agarose gel electrophoresis to ensure that no degradation had occurred (data not shown).

Reverse Transcription and PCR Amplification

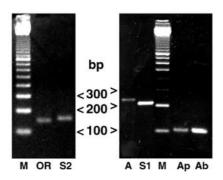
Reverse transcription and PCR amplifications were performed with the GeneAmp® RNA PCR kit (Perkin-Elmer, Barcelona, Spain). First, cDNAs were synthesized by reverse transcription from 0.5 μ g of total RNA. This reaction was performed in PCR buffer (10 mM Tris-HCl, 50 mM KCl, pH 8.3), containing 5 mM MgCl₂, 1 mM of each dNTP, 20 U of RNase inhibitor, 50 U of Moloney murine leukemia virus reverse transcriptase, and 2.5 μ M of random hexamers in a final volume of 20 μ l. The mixture was incubated for 5 min at 25°C, 15 min at 42°C, 5 min at 98°C, and 5 min at 10°C.

The primers for PCR amplifications were chosen from coding regions of each gene that exhibited poor homology to related GPCRs or other known human genes, and they were designed to produce amplicons of different sizes (Table 2 and Figure 1). Considering these restrictions, the length and GC content of forward and reverse primers was maintained as close as possible within each primer set. PCR amplifications were carried out in PCR buffer containing

Table 2 Primer Sequences for PCR Amplification of α_{2a} -Adrenoceptor, 5-HT_{1a}, 5-HT_{2a}, μ -Opioid Receptor, and β -Actin cDNAs

Template	Sequence	L	%GC	Product size (bp)
α_{2a} -Adrenoceptor	Forward: 5' GCAGAGGCCGAACCGCTGCCCACCCAGCTC 3'	30	73	
	Reverse: 5' AGCCGGCCGAGCCGCGCTGCGAGAT 3'	25	76	103
5-HT _{IA} receptor	Forward: 5'ACCCCGGAAGACCGCTCGGACCCCGACGCA 3'	30	73	
	Reverse: 5' CTGCGGGGGGGGAGATGCTCCATGGCGGGT 3'	30	73	219
5-HT _{2A} receptor	Forward: 5' CAGGACGATTCGAAGGTCTTTAAGGAGGGG 3'	30	53	
	Reverse: 5' CCGTGTGCCAAGATCACTTACACAAAGT 3'	30	47	180
μ -Opioid receptor	Forward: 5' CCAGAAACTACGTTCCAGACTGTT 3'	24	46	
	Reverse: 5' AGTGGAGTTTTGTTGCTCAAT 3'	21	38	168
β -Actin	Forward: 5' ATCGTGGGCCGCCCTAGGCA 3'	20	70	
	Reverse: 5' GGGGAGACTTGGGATTCCGGT 3'	21	62	240

The length (L) and GC content (%GC) of each primer, as well as the length of the PCR product (base pairs) for each set of primers, are indicated. For other details see text.



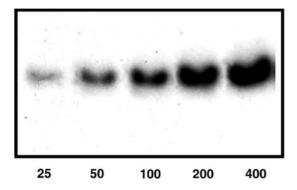


Figure I Upper panel: Agarose gel electrophoresis of RT-PCR amplification products of the human brain α_{2A} -adrenoceptor (Ab), 5- HT_{1A} (S1), 5- HT_{2A} (S2), and μ -opioid (OR) receptors, and β -actin (A). The human platelet α_{2A} -adrenoceptor (Ap) was also amplified as a positive control since this is the only α_2 -adrenoceptor subtype expressed in platelets. Negative controls (no RNA or cDNA and no reverse transcriptase) did not yield PCR products (data not shown). A 100-base pair ladder (M) was used to determine product sizes. In addition to the correct size, PCR products were sequenced to confirm their identity. For other details see the Subjects and Methods section. Lower panel: RT-PCR amplifications corresponding to a standard curve for the α_{2A} -adrenoceptor. The amount of total RNA and the corresponding IOD values for the bands shown were: 25 ng and 2628 IOD units; 50 ng and 8754 IOD units; 100 ng and 20894 IOD units; 200 ng and 42115 units; and 400 ng and 73726 IOD

2 mM MgCl₂, 1 mM of each dNTP, 1 nmol of [³²P]dCTP (3 μl of a 1:10 dilution of 3000 Ci/mmol and 10 mCi/ml, Amersham Pharmacia Biotech, Barcelona, Spain), 2.5 U of Ampli Taq DNA polymerase, and 0.15 µM of each (forward and reverse) primer, in a total volume of 100 µl. Other experimental conditions (volume of reverse transcription reaction used for PCRs, annealing temperatures, and number of cycles) were determined empirically for each mRNA species. The conditions were optimized to obtain a linear relation between the initial amount of RNA and the integrated optical density (IOD) of the signal obtained for each product (see below). For α_{2A} -adrenoceptors, 5-HT_{1A}, 5-HT_{2A}, and μ -opioid receptors, 4 μ l of reverse-transcription reaction medium was used for PCR amplifications (corresponding to 100 ng of total RNA in the reverse transcription reaction). For β -actin 0.3 μ l (3 μ l from a 1:10 dilution, corresponding to 7.5 ng of total RNA) was used. For quantitative evaluations, standard curves of five PCR reactions containing various amounts of cDNA template from one control subject were prepared. For standard



curves to quantify α_{2A} -adrenoceptors, 5-HT_{1A}, 5-HT_{2A}, and μ -opioid receptors, 1–16 μ l of reverse transcription reaction medium was used (25, 50, 100, 200, and 400 ng of total RNA). For standard curves of β -actin 0.1–1 μ l (1–10 μ l from a 1:10 dilution, corresponding to 2.5, 5, 7.5, 15, and 25 ng of total RNA) was used. Before each reaction, the cDNA templates (from the reverse transcription reaction) were diluted in water to a final volume of 20 µl, heated to 80°C, and added to tubes containing the other PCR reagents (80 μl), previously incubated for 2 min at 80°C to avoid nonspecific annealing ('hot start'). Then, following a 2-min denaturalization step at 94°C, thermal cycling commenced. The thermal cycles consisted of two or three steps at the following temperatures and times: 94°C for 1 min, 62°C for 1 min, and 72 $^{\circ}$ C for 20 s (30 cycles) for α_{2A} -adrenoceptors and 5-HT_{1A} receptors; 94°C for 1 min, 51°C for 1 min, and 72°C for 30 s (31 cycles) for μ -opioid receptors and β -actin; 94°C for 1 min and 65°C for 1 min (30 cycles) for 5-HT_{2A} receptors. After thermal cycling, the samples were maintained at 72°C for 7 min and the PCR products were analyzed as indicated below.

Detection and Quantification of mRNA Expressions

Preliminary series of RT-PCRs were carried out to ascertain the nature of the amplified PCR products. For these series, the methods used were the same as above, except that the number of thermal cycles was 35-36, instead of 30-31, and [³²P]dCTP was not included in the reaction mix. After PCR, $10\,\mu l$ of reaction mixture was combined with $2\,\mu l$ of electrophoresis loading buffer (0.25% bromophenol blue, 0.25% xylene cyanol FF, 30% glycerol) and separated on 2% agarose gels in TAE buffer (40 mM Tris-Acetate, 1 mM EDTA) containing SYBR Green I (1:10000 dilution; Molecular Probes, Barcelona, Spain). Under these experimental conditions, each RT-PCR produced a unique band of the expected size (Figure 1, Table 2). These PCR products were further analyzed by DNA sequencing of both the sense and antisense DNA strands, which confirmed the origin of the amplified DNA.

PCR reactions in which the amplified products were labelled with [32P]dCTP (since only 30-31 cycles were used to fall into the linear part of the PCR-kinetics sigmoid curve) produced signals detectable by autoradiography, whose intensities were proportional to the initial RNA concentration used for the RT-PCR (Figure 1). To detect the PCR products, 10 µl of the PCR mixture was combined with 2 μl of electrophoresis loading buffer and the products were separated by electrophoresis on 15-well 7% polyacrylamide gels $(6 \times 8 \text{ cm}^2, 0.75 \text{ cm thickness})$ in TBE buffer (45 mM)Tris-Borate, 1 mM EDTA). The evaluation of mRNA expression from each subject and for each receptor was performed in three independent RT-PCR experiments. From each experiment, four test samples and four control samples were loaded along with the standards (five samples of known concentration) on the same gel, for a total of 12 measurements per subject and mRNA species. After electrophoresis, gels were dried, covered with a plastic wrapping, and exposed to autoradiographic film (Hyperfilm, Amersham) with an intensifying screen at -80° C for between 15 min and 4 h. The IOD values of autoradiography bands were determined by image analysis using the



program SigmaGel (SPSS Science, Erkrath, Germany). To quantify the mRNA levels, the IOD values from the test samples were compared to those from the standards as described previously (Escribá et al, 1994). To avoid artifacts, mRNA expression for each receptor was corrected by the expression of β -actin, which was used as internal control, being determined from the same cDNA sample used to measure receptor expressions, as indicated above. Therefore, the mRNA expression for each receptor was calculated as RE = $[(R_R/R_T)/(A_R/A_T)] \times 100$, where R_R is the calculated mRNA content for a given receptor, obtained by interpolation of the autoradiography band IOD value into the standard control curve; R_T is the total amount of mRNA present in the PCR, calculated by spectrophotometry; and A_R and A_T are equivalent to R_R and R_T , but for β -actin.

Statistics

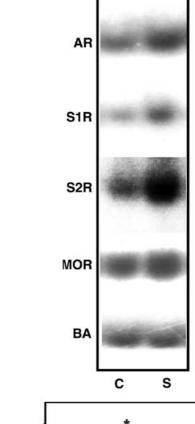
The results were finally expressed as the means \pm SEM. One-sample *t*-test (identical in its implementation to the paired *t*-test) was used for statistical evaluations. Pearson correlation coefficients were calculated to test for possible associations among variables. The level of significance was $p \le 0.05$. All tests were two-tailed.

RESULTS

RT-PCR has been used to determine gene expression in many types of tissue, including brain tissue from both humans and animals (Cao et al, 2001; Li and Wang, 2000; Newman et al, 1988). Here, we have used this method to quantify the mRNA levels of various GPCRs in post-mortem brain tissue samples from suicide victims and control subjects. In contrast to other similar experimental approaches, such as 'real-time RT-PCR', the technique employed here separates undesired nonspecific reaction products from those amplified specifically, thereby eliminating a potential artifact from the analysis. However, under our experimental conditions, no nonspecific products were detected at all (Figure 1). In addition, the specificity of these primers was further confirmed by sequencing the reaction products generated on both strands.

GPCR mRNA Expression in the Brains of Suicide Victims

Major depression has been associated with alterations in the density of $\alpha_{2\mathrm{A}}$ -adrenoceptors, 5-HT $_{1\mathrm{A}}$, 5-HT $_{2\mathrm{A}}$, and μ -opioid receptor proteins. Consistent with these findings, we found that the level of $\alpha_{2\mathrm{A}}$ -adrenoceptor mRNA augmented in the prefrontal cortex of suicide victims (42.5 \pm 10.5% over cotrol, p=0.03; Figure 2). Similarly, quantitative analyses demonstrated marked and significant increases in the expression of mRNA for the 5-HT $_{1\mathrm{A}}$, 5-HT $_{2\mathrm{A}}$ receptors (131 \pm 33% over control, p=0.02 and 65 \pm 26% over control, p=0.04, respectively; Figure 2). Furthermore, expression of the μ -opioid receptor was also augmented in suicide subjects (89 \pm 36% over control, p=0.03; Figure 2). A comparison between the increase in the levels of receptor expression in the frontal cortex of suicide victims with a known (n=9) or unknown (n=3) history of major



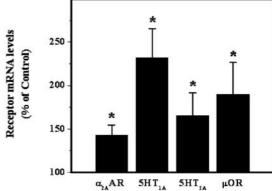


Figure 2 GPCRs expression in the human brain. Upper panel: Comparative expression of α_{2A} -adrenoceptors (AR), 5-HT_{1A} (S1R), 5-HT_{2A} (S2R), and μ -opioid receptors (MOR) in the brain of a depressed suicide victim (S) and an age-, sex-, and post-mortem-delay-matched control (C). In both cases, 100 ng of total RNA was used for RT-PCR. The expression of β-actin (BA) is also shown. Lower panel: Relative expressions of GPCRs (α_{2A} -AR, α_{2A} -adrenoceptors; 5HT_{1A}, 5-HT_{1A} receptors, 5HT_{2A}, 5-HT_{2A} receptors; μ OR, μ -opioid receptors) in the brains of suicide victims. Data are the percentage values (mean ± SEM) with respect to the matched control group (100%). *p < 0.05.

depression did not show any significant differences (2–12%, p>0.05). Nor were significant changes observed in the relative levels of β -actin in the suicide group (92 \pm 16%) with respect to the control group (99 \pm 14%). Thus, in this study, the levels of receptor expression in each brain sample were corrected for the relative levels of β -actin in the same brain sample to avoid possible artifacts originated by RNA degradation, differences in RNA concentration measurements, etc.

Correlation between the Levels Of α_{2a} -adrenoceptor mRNA and Protein in the Human Brain

In previous studies, we have used other biochemical and immunological approaches to investigate α_2 -adrenoceptor protein in suicide and depressed subjects (Callado et al, 1998; García-Sevilla et al, 1999). However, due to differences in the regulation of the translational and post-translational machinery, changes in the cellular levels of an individual mRNA species and its corresponding protein are not always equivalent. For this reason, we analyzed the relationship between α_{2A} -adrenoceptor mRNA and protein levels. A significant correlation (r = 0.79; p < 0.05) was found between the expression of α_{2A} -adrenoceptor mRNA when compared with the corresponding levels of protein (measured by immunoblotting). The data corresponding to adrenoceptor protein were measured previously in the same brain area of those subjects in which both molecular species were measured (Brodmanńs area 9, n = 9; Figure 3) (García-Sevilla *et al*, 1999).

Relationships Between the mRNA Levels of GPCRs

The GPCRs here studied share signaling entities and expression regulatory DNA sequences (see below), so that we investigated possible relationships in the expressions of these receptors. In the human prefrontal cortex, expression of α_{2A} -adrenoceptors was closely correlated with that of 5- HT_{1A} receptors (r = 0.6, p = 0.014, n = 19; Figure 4). The expression of the latter also showed a significant correlation with the expression of 5-HT_{2A} receptors (r = 0.65, p = 0.007, n = 18; Figure 4). In addition, 5-HT_{2A} receptor expression showed a trend for correlation with the expression of both the α_{2A} -adrenoceptors (r = 0.38, p = 0.12, n = 18) and

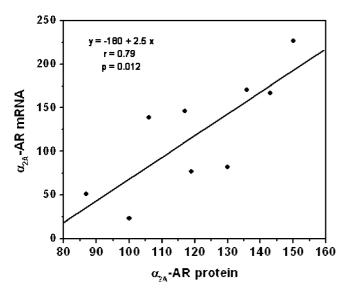
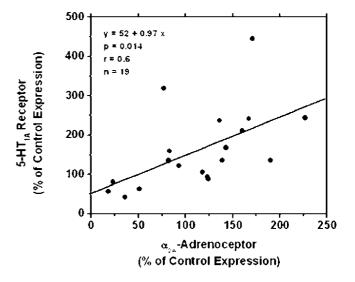


Figure 3 Correlation between α_{2A} -adrenoceptor density (protein levels) and expression (mRNA levels) measured in the same brain area (Brodmann's area 9) of the same subjects. The density of α_{2A} adrenoceptors was determined by immunoblotting, as previously described (García-Sevilla et al, 1999). Data are expressed as the percentage of immunoreactivity or expression with respect to control samples. Each point represents a different subject and the solid line represents the regression of the correlation. The data were best described by the expression $y = -180 + 2.5 \times (r = 0.79, n = 9, p = 0.012).$



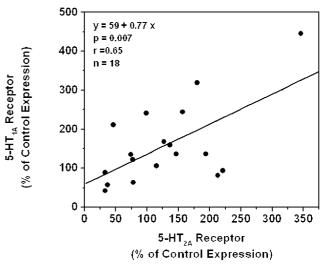


Figure 4 Correlations between the expression of α_{2A} -adrenoceptors and 5-HT $_{1A}$ receptors (upper panel), and 5-HT $_{1A}$ and 5-HT $_{2A}$ receptors (lower panel) in the brains of the same subjects (controls and suicides). Each point represents a different subject and the solid lines represent the regression of the correlations. Both axes are expressed as percentages of receptor mRNA levels with respect to a control subject (100%). Significant correlations were found between the two variables in both cases. The data were best described by the expressions: α_{2A} -adrenoceptors vs 5-HT_{IA} receptors, $y = 52 + 0.97 \times (r = 0.6, n = 19, p = 0.014)$ and 5-HT_{IA} vs 5-HT_{2A} receptors, $y = 59 + 0.77 \times (r = 0.65, n = 18, p = 0.007)$. Correlations between other variables were not significant (see the Results section).

 μ -opioid receptors (r = 0.43, p = 0.09, n = 17). Similarly, the expression of 5-HT_{1A} receptors showed a trend for correlation with that of the μ -opioid receptors (r = 0.42, p = 0.1, n = 17). α_{2A} -Adrenoceptor expression did not appear to be related with that of μ -opioid receptors (r = 0.18, p = 0.47, n = 17).

DISCUSSION

To our knowledge, this is the first time that alterations in the expression of α_{2A} -adrenoceptor, serotonin (1A and 2A subtypes) receptors, and the μ -opioid receptor have been





reported in the frontal cortex of the same suicide victims. In addition, there is no information about adrenoceptor and opioid receptor mRNA levels and very few reports about the expression of 5-HT receptors and associated signaling elements in suicides (Arango et al, 2001; Pandey et al, 2002). This study was designed to evaluate the molecular alterations in the post-mortem brain of subjects with mood disorders who had committed suicide. Depression is the most important risk factor for suicide and, therefore, a common approach to study neurochemical alterations in mood disorders is the analysis of brain samples from suicide victims (García-Sevilla et al, 1996a). Indeed, no significant differences were found between GPCR expression in the brains of suicide victims with known (n = 9) and unknown (n=3) history of depression. Therefore, the conclusions drawn from the present results might be relevant to both suicide and depression. As the molecular bases of suicide and depression are not fully understood, we studied the expression of various GPCRs associated with these human pathologies. In these studies, a number of issues were raised and answered.

Alterations in α_{2a} -AR Expression in the Brains of Suicides

Presynaptic inhibitory α_{2A} -adrenoceptors control neuronal norepinephrine and 5-HT release. Previous studies, using specific antibodies, have demonstrated that the density of α_{2A} -adrenoceptor is increased in suicide subjects with major depression (García-Sevilla et al, 1999). In addition, other studies using radioligand agonists revealed an elevation in the density of α_{2A} -adrenoceptor (high-affinity state) in the post-mortem brains of suicide victims (Callado *et al*, 1998; De Paermentier et al, 1997; González et al, 1994; Meana et al, 1992; Meana and García-Sevilla, 1987). In addition, higher levels of G_i proteins (García-Sevilla et al, 1996b, 1997, 1999) and lower levels of adenylyl cyclase activity (Reiach et al, 1999) were reported in the platelets of depressed subjects and post-mortem brains of suicide victims who previously suffered from depression. Finally, the activity of α_{2A} adrenoceptors has recently been found to be enhanced in the brains of depressed suicide victims, as determined by [35S]GTPyS binding (González-Maeso et al, 2002). This is in agreement with studies demonstrating increased receptor and Gi protein densities. These results further support the hypothesis that adrenoceptor supersensitivity explains in part the molecular alterations related to suicide and depression (García-Sevilla et al, 1986). Thus, increases of α_2 -adrenoceptor and Gi protein density/activity, and decreases of adenylyl cyclase density/activity would be involved with reductions in cytosolic levels of cAMP. Moreover, antidepressant drug treatment has been shown to decrease the levels of α_{2A} -adrenoceptors and Gi proteins, further implicating these receptors in the pathophysiology, and thus, in the therapy of suicide, and depression (De Paermentier et al, 1997; García-Sevilla et al, 1986; Piletz et al, 1996).

All these data indicate that the α_{2A} -adrenoceptor signal pathway is enhanced at each molecular step, from receptor to second messenger in the brains of suicide victims. In this context, significant increases in α_{2A} -adrenoceptor mRNA levels were found in the post-mortem brains of suicides.

This demonstrates that increases in the receptor protein detected by immunoblotting and radioligand binding studies originate from an increase in adrenoceptor transcription and not by other causes, such as altered receptor turnover. In addition to protein studies, it is advisable to investigate gene expression, since discrepancies between protein and mRNA levels in signaling proteins have been previously reported and associated with post-translational regulatory processes (Young et al, 1987; Busquets et al, 1997; Ventayol et al, 1997).

Besides these novel alterations in the expression of these GPCRs in the brains of suicide victims, we also studied the correlation between mRNA and protein levels in the brains of the same subject and brain area. A positive correlation was found between both molecular entities (α_{2A} -adrenoceptor protein and mRNA). This result indicates that posttranslational mechanisms and proteolytic degradation do not play important roles in the regulation of this receptor in vivo, as previously suggested from studies of the adrenoceptor stability (Ozaita et al, 1999).

Altered 5-HT Receptor Expression in the Brains of **Suicide Victims**

Brain 5-HT receptors have also been implicated in the pathophysiology of suicide. A large number of studies have demonstrated alterations in the abundance and/or function of these receptor proteins in depressive/suicide subjects, but there is a lack of information regarding the mRNA levels of these receptors in the brains of suicide victims. The levels of the 5-HT_{2A} receptor protein have been consistently found to be increased in depressed subjects, (Mendelson, 2000) and increased coupling and activity of 5-HT_{2A} receptors has been demonstrated in depressed patients (Serres et al, 1999). As a consequence, this receptor has been proposed as a marker for depression/suicide (Mendelson, 2000; Pandey et al, 1995). In this work, we found a significant degree of 5-HT_{2A} receptor overexpression in the frontal cortex of depressed suicides, in agreement with data recently published (Pandey et al, 2002). In addition, positron emission tomography studies suggest that depressive and suicidal states are associated with increases of 5-HT₂ receptors and low levels of the 5-HT metabolite 5hydroxyindoleacetic acid, which suggest reduced neurotransmitter concentrations (Meyer et al, 1999, 2003). Thus, as for α_{2A} -adrenoceptors, these increases in 5-HT_{2A} receptor density and function could be attributed to an increased rate of transcription. On the other hand, a significant correlation between the expression of 5-HT_{2A} receptors and α_{2A} -adrenoceptors suggests that they might share common regulatory elements and further supports their involvement in the pathophysiology of suicide. However, it is not clear what the origin of the common regulation of these two presynaptic receptors might be, and whether heterologous regulation, cross-talk mechanisms, or common transcription regulatory processes could be involved in this phenomenon (see below).

In the present study, it has also been shown that the expression of 5-HT_{1A} receptors increases in the brain of suicide victims. This result is in agreement with a recent report, showing that the promoter region of the 5-HT_{1A} gene contains a polymorphism that accounts for increased

receptor expression in association with depression and suicide (Lemonde et al, 2003). Studies on the alterations of 5-HT_{1A} receptors in suicide/depressed subjects have generated certain controversy, since both increases and decreases in receptor protein abundance have been reported. It has been suggested that impaired receptor sensitivity (Cowen et al, 1994) or altered signaling (Lesch et al, 1990) could account for the apparent decreases in receptor abundance, although post-transcriptional regulation and protein degradation (Young et al, 1987; Busquets et al, 1997; Ventayol et al, 1997) could also be involved in the differences observed between receptor mRNA and protein levels. In fact, a functional impairment of 5-HT_{1A}-associated signaling is originated by alteration of downstream transduction proteins in the brains of suicides (Hsiung et al, 2003). Moreover, a lower serotonergic activity associated with suicidal behavior has been also shown through positron emission tomography studies in depressed subjects and radioligand binding studies in post-mortem brain of suicides (Oquendo et al, 2003; Stockmeier et al, 1998). On the other hand, many antidepressant drug treatments downregulate 5-HT_{1A} and 5-HT_{2A} receptors, supporting the idea that increases in the activity of both receptors are associated with the development of suicidal/depressive behaviors (Stahl, 1994). In the present study, a significant correlation between the expression of 5-HT_{1A} receptors and 5-HT_{2A} receptors was also found. Gene expression is modulated by proteins acting at DNA regulatory sequences. In this context, it has been shown that SP1- and cAMP responsive (CRE)-related DNA sequences regulate 5-HT_{1A}, 5-HT_{2A}, and α_{2A} -adrenoceptor expression (Handy and Gavras, 1996; Tamotsu et al, 1995; Meijer et al, 2000; Zhu et al, 1995). Thus, regulation by common transcription activators and repressors could be involved in the coordinated changes in α_2 -adrenoceptor and 5-HT receptor mRNA levels that we observed.

Altered Expression of μ -Opioid Receptors in the Brains of Suicide Victims

Most of the biochemical studies on neuroreceptor alterations in suicide brain tissue have been focused on adrenoceptors and serotonin receptors. However, clinical and biochemical evidence of alterations of the μ -opioid receptor in suicide also exists. In line with the present results, it was previously shown that the density of μ -opioid receptor sites is augmented in the post-mortem brain of depressed suicides (Gross-Isseroff et al, 1990; Gabilondo et al, 1995). Moreover, treatments with antidepressant drugs induce decreases in the receptor (and associated neuropeptide) levels (Hamon et al, 1987). Here we have shown that the expression of the μ -opioid receptor is significantly and markedly increased in the frontal cortex of suicide victims, supporting the involvement of this receptor in the pathophysiology of suicide. In this sense, it has been recently shown that stress, related to certain suicidal behaviors, induces increases in μ -opioid receptor mRNA expression in the rat brain (Yamamoto et al, 2003). In addition, positron emission tomography studies in humans demonstrate the in vivo involvement of these receptors in mood and behavior, further suggesting their potential implication in suicide behaviors (Zubieta et al, 2003). Recent [35 S]GTP γ S binding studies, however, indicate that the activity of the μ -opioid receptor signaling is not altered in the brains of suicide victims (González-Maeso *et al*, 2002), despite the higher mRNA and protein levels found in this and other reports. The same reasons discussed above for 5-HT receptors might be involved in this phenomenon. On the other hand, no correlation was found between the expression of μ -opioid receptors and those of 5-HT receptors and α_{2A} -adrenoceptors, suggesting that some divergent regulatory mechanisms control the expressions of these receptors.

Is RT-PCR a Suitable Technique to Measure Receptor Expression in Human Post-mortem Brain Samples?

A number of techniques have been used to evaluate the rate of gene transcription in cells and tissues. However, more recently PCR-based methods have been used to detect smaller amounts of mRNA in human samples (Cao et al, 2001; Li and Wang, 2000; Newman et al, 1988). In this study, we have shown that RT-PCR is suitable for the quantification of certain mRNA species in post-mortem human brain tissue. However, when performing such analyses, several factors should be taken into account. Firstly, mRNA degradation after death can lead to the fragmentation of RNA species (Barton et al, 1993; Leonard et al, 1993). The longer the time spent by the corpses at high outdoor temperatures, the more severe mRNA degradation is. RNA fragmentation makes it difficult to detect gene expression by Northern blot or other hybridization techniques. However, fragmented mRNA may still be a good substrate for RT-PCR, since amplicons usually correspond to short regions of mRNA (100-300 bases), whose length might be smaller that that of the RNA fragments. Additionally, cuts within these regions can still yield amplification products. Furthermore, mRNA degradation (and other possible artifacts) can be corrected for by measuring the expression of a housekeeping gene, such as β -actin, and expressing the levels of expression of the mRNA of interest relative to the expression of such genes, as performed here.

The effect of other variables (age, sex, post-mortem delay, drugs, etc.) was also evaluated. For the receptors here studied and under the experimental conditions used, only elevated post-mortem delay periods appeared to induce decreases in the amount of mRNA in the human post-mortem brain. However, when receptor mRNA content was corrected by β -actin mRNA content, no difference in the relative content of mRNA was detected among subjects with different post-mortem delays. This result further supports the convenience to use an internal reference rather that an external cDNA to evaluate the overall experimental process. In addition, the very small difference between the post-mortem delay values of the study and control groups (only about 40 min) discounted possible influences of this variable on the final result obtained.

One of the advantages of using RT-PCR for this type of analysis is that it is very sensitive, and thus, small amounts of template mRNA can be detected and measured. Moreover, mRNA levels for a given receptor are not masked by other mRNA species, whereas when measuring GPCR protein levels through functional properties (eg agonist



radioligand binding), the behavior of these receptors can be modified by G protein coupling, receptor phosphorylation, internalization, etc. (which may result in alterations in the apparent protein levels). Finally, the close correlation between template (mRNA/cDNA) concentrations and the signal detected (IOD values), as well as the high signal-tonoise ratio obtained demonstrated that RT-PCR is a suitable technique for the study of receptor expression in postmortem human brain samples. In contrast, other techniques are not so advantageous and often give qualitative results difficult to quantify.

ACKNOWLEDGEMENTS

This work was supported by Grants BFI2000-0306 (JAGS), SAF2001-0839, SAF 2003-00232 and FIS00/1029 (PVE). AO was supported by a fellowship from the Basque Country.

REFERENCES

- Arango V, Underwood MD, Boldrini M, Tamir H, Kassir SA, Hsiung S *et al* (2001). Serotonin 1A receptors, serotonin transporter binding and serotonin transporter mRNA expression in the brainstem of depressed suicide victims. *Neuropsychopharmacology* **25**: 892–903.
- Barton AJL, Pearson RC, Najlerahim A, Harrison PJ (1993). Preand postmortem influences on brain RNA. J Neurochem 61: 1-11.
- Busquets X, Ventayol P, García-Sevilla JA (1997). Naloxone-precipitated withdrawal in morphine-dependent rats increases the expression of α_{2A} -adrenoceptor mRNA in brain. *Mol Brain Res* 45: 154–158.
- Callado LF, Meana JJ, Grijalva B, Pazos A, Sastre M, García-Sevilla JA (1998). Selective increase of alpha-2A-adrenoceptor agonist binding sites in brains of depressed suicide victims. *J Neurochem* **70**: 1114–1123.
- Cao Z, Wang Y, Reid EA, McShepard G, Kemp M, Newkirk RF et al (2001). The quantitative distribution of a putative PKC epsilon mRNA in Limulus central nervous system by modified competitive RT-PCR. J Neurosci Methods 105: 193–199.
- Chomczynski P, Sacchi N (1987). Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* **162**: 156–159.
- Chuang TT, Lacovelli L, Sallese M, De Blasi A (1996). G protein-coupled receptors: heterologous regulation of homologous desensitization and its implications. *Trends Pharmacol Sci* 17: 416–421.
- Cowen PJ, Power AC, Ware CJ, Anderson IM (1994). 5-HT1A receptor sensitivity in major depression. A neuroendocrine study with buspirone. *Br J Psychiatry* **164**: 372–379.
- De Paermentier F, Mauger JM, Lowther S, Crompton MR, Katona CLE, Horton RW (1997). Brain α_{2A} -adrenoceptors in depressed suicides. *Brain Res* 757: 60–68.
- Escribá PV, Sastre M, García-Sevilla JA (1994). Increased density of guanine nucleotide-binding proteins in the postmortem brains of heroin addicts. *Arch Gen Psychiatry* **51**: 494–501.
- Esteban S, Lladó J, Sastre-Coll A, García-Sevilla JA (1999). Activation and desensitization by cyclic antidepressant drugs of α_2 -autoreceptors, α_2 -heteroreceptors and 5-HT_{1A}-autoreceptors regulating monoamine synthesis in the rat brain *in vivo*. *Naunyn-Schmiedeberg's Arch Pharmacol* **360**: 135–143.
- Gabilondo AM, Meana JJ, García-Sevilla JA (1995). Increased density of μ-opioid receptors in the postmortem brain of suicide victims. *Brain Res* 682: 245–250.

- García-Sevilla JA, Escribá PV, Busquets X, Walzer C, Guimón J (1996b). Platelet imidazoline receptors and regulatory G proteins in patients with major depression. *Neruoreport* 8: 169–172.
- García-Sevilla JA, Escribá PV, Ozaita A, La Harpe R, Walzer C, Eytan A *et al* (1999). Up-regulation of immunolabeled α_{2A} -adrenoceptors, Gi coupling proteins and regulatory receptor kinases in the prefrontal cortex of depressed suicides. *J Neurochem* 72: 282–291.
- García-Sevilla JA, Escribá PV, Sastre M, Walzer C, Busquets X, Jaquet G et al (1996a). Immunodetection and quantitation of imidazoline receptor proteins in platelets of patients with major depression and in brains of suicide victims. Arch Gen Psychiatry 53: 803–810.
- García-Sevilla JA, Guimón J, García-Vallejo P, Fuster MJ (1986). Biochemical and functional evidence of supersensitive platelet α_2 -adrenoceptors in major affective disorder: effect of long-term lithium carbonate treatment. *Arch Gen Psychiatry* **43**: 51–57.
- García-Sevilla JA, Walzer C, Busquets X, Escribá PV, Balant L, Guimon J (1997). Density of guanine nucleotide-binding proteins in platelets of patients with major depression: increased abundance of the Gαi₂ subunit and down-regulation by antidepressant drug treatment. *Biol Psychiatry* 42: 704–712.
- González AM, Pascual J, Meana JJ, Barturen F, del Arco C, Pazos A et al (1994). Autoradiographic demonstration of increased α_2 -adrenoceptor agonist binding sites in the hippocampus and frontal cortex of depressed suicide victims. *J Neurochem* 63: 256–265.
- González-Maeso J, Rodríguez-Puertas R, Meana JJ, García-Sevilla JA, Guimón J (2002). Neurotransmitter receptor-mediated activation of G-proteins in brains of suicide victims with mood disorders: selective supersensitivity of α_{2A} -adrenoceptors. *Mol Psychiatry* 7: 755–767.
- Gross-Isseroff R, Dillon KA, Israeli M, Biegon A (1990). Regionally selective increases in μ opioid receptor density in the brains of suicide victims. *Brain Res* **530**: 312–316.
- Hamon M, Gozlan H, Bourgoin S, Benoliel JJ, Mauborgne A, Taquet H *et al* (1987). Opioid receptors and neuropeptides in the CNS in rats treated chronically with amoxapine and amitriptyline. *Neuropharmacology* **26**: 531–539.
- Handy DE, Gavras H (1996). Evidence for cell-specific regulation of transcription of the rat α_{2A} -adrenergic receptor gene. *Hypertension* 27: 1018–1024.
- Hsiung SC, Adlesberg M, Arango V, Mann JJ, Tamir H, Liu KP (2003). Attenuated 5-HT_{1A} receptor signaling in brains of suicide victims: involvement of adenylyl cyclase, phosphatidylinositol 3-kinase, Akt and mitogen-activated protein kinase. *J Neurochem* 87: 182–194.
- Invernizzi RW, Parini S, Sacchetti G, Fracasso C, Caccia S, Annoni K *et al* (2001). Chronic treatment with reboxetine by osmotic pumps facilitates its effect on extracellular noradrenaline and may desensitize α_2 -adrenoceptors in the prefrontal cortex. *Br J Pharmacol* 132: 183–188.
- Lemonde S, Turecki G, Bakish D, Du L, Hrdina PD, Bown CD *et al* (2003). Impaired regression at a 5-hydroxytryptamine 1A receptor gene polymorphism associated with major depression and suicide. *J Neurosci* 23: 8788–8799.
- Leonard S, Logel J, Luthman D, Casanova M, Kirch D (1993). Freedman R. Biological stability of mRNA isolated from human postmortem brain collections. *Biol Psychiatry* 33: 456–466.
- Lesch KP, Mayer S, Disselkamp-Tietze J, Hoh A, Wiesmann M, Osterheider M *et al* (1990). 5-HT1A receptor responsivity in unipolar depression. Evaluation of ipsapirone-induced ACTH and cortisol secretion in patients and controls. *Biol Psychiatry* **28**: 620–628.
- Li X, Wang X (2000). Aplication of real-time polymerase chain reaction for the quantitation of interleukin-1beta mRNA upregulation in brain ischemic tolerance. *Brain Res Protocols* 5: 211–217.

- Mann JJ (1999). Role of the serotonergic system in the pathogenesis of major depression and suicidal behavior. Neuropsychopharmacology 25: 99S-105S.
- Meana JJ, Barturen F, García-Sevilla JA (1992). α_2 -Adrenoceptors in the brain of suicide victims: increased receptor density associated with major depression. Biol Psychiatry 31: 471-490.
- Meana JJ, García-Sevilla JA (1987). Increased α_2 -adrenoceptor density in the frontal cortex of depressed suicide victims. J Neural Transm 70: 377-381.
- Meijer OC, Williamson A, Dallman MF, Pearce D (2000). Transcriptional repression of 5-HT_{1A} promoter by corticosterone via mineralocorticoid receptors depends on the cellular context. J Neuroendocrinol 12: 245-254.
- Mendelson SD (2000). The current status of the platelet 5-HT_{2A} receptor in depression. J Affect Disord 57: 13-24.
- Meyer JH, Kapur S, Houle H, DaSilva J, Owczarek B, Brown GM et al (1999). Prefrontal cortex 5-HT2 receptors in depression: an [18F]setoperone PET imaging study. Am J Psychiatry 156: 1029-1034.
- Meyer JH, McMain S, Kennedy SH, Korman L, Brown GM, DaSilva JN et al (2003). Dysfunctional attitudes and 5-HT2 receptors during depression and self-harm. Am J Psychiatry 160: 90-99.
- Neumaier JF, Root DC, Hamblin MW (1996). Chronic fluoxetine reduces serotonin transporter mRNA and 5-HT_{1B} mRNA in a sequential manner in the rat dorsal raphe nucleus. Neuropsychopharmacology 15: 515-522.
- Newman PJ, Gorsky J, White II GC, Gidwitz S, Cretney CJ, Aster RH (1988). Enzymatic amplification of platelet-specific messenger RNA using the polymerase chain reaction. J Clin Invest 82:
- Nordstrom P, Samuelsson M, Asberg M, Traskman-Bendz L, Aberg-Wistedt A, Nordin C et al (1994). CSF 5-HIAA predicts suicide risk after attempted suicide. Suicide Life Threat Behav
- Oquendo MA, Placidi GP, Malone KH, Campbell C, Keilp J, Brodsky B et al (2003). Positron emission tomography of regional brain metabolic responses to a serotonergic challenge and lethality of suicide attempts in major depression. Arch Gen Psychiatry 60: 14-22.
- Ordway GA, Widdowson PS, Smith KS, Halaris A (1994). Agonist binding to α_2 -adrenoceptors is elevated in the locus coeruleus from victims of suicide. J Neurochem 63: 617-624.
- Ozaita A, Escribá PV, García-Sevilla JA (1999). The alkylating agent EEDQ facilitates protease-mediated degradation of the human brain α_2 -adrenoceptor as revealed by a sequence-specific antibody. Neurosci Lett 263: 105-108.
- Pandey GN, Dwivedi Y, Rizavi HS, Ren X, Pandey SC, Pesold C et al (2002). Higher expression of serotonin 5-HT_{2A} receptors in the postmortem brains of teenage suicide victims. Am J Psychiatry **159**: 419–429.
- Pandey GN, Pandey SC, Dwivedi Y, Sharma RP, Janicak PG, Davis JM (1995). Platelet serotonin-2A receptors: a potential biological marker for suicidal behavior. Am J Psychiatry 152: 850-855.
- Piletz JE, Halaris A, Nelson J, Qu Y, Bari M (1996). Platelet I₁-imidazoline binding sites are elevated in depression but not generalized anxiety disorder. J Psychiatry Res 30: 147-168.
- Reiach JS, Li PP, Warsh JJ, Kish SJ, Young LT (1999). Reduced adenylyl cyclase immunolabeling and activity in postmortem temporal cortex of depressed suicide victims. J Affect Disord 56:
- Ressler KJ, Nemeroff CB (1999). Role of norepinephrine in the pathophysiology and treatment of mood disorders. Biol Psychiatry 46: 1219-1233.
- Roy A, Agren H, Pickar D, Linnoila M, Doran AR, Cutler NR et al (1986). Reduced CSF concentrations of homovanillic acid and homovanillic acid to 5-hydroxyindolacetic acid ratios in

- depressed patients: relationship to suicidal behavior and dexamethasone nonsuppression. Am J Psychiatry 143: 1539-1545.
- Sargent PA, Kjaer KH, Bench CJ, Rabiner EA, Messa C, Mayer J et al (2000). Brain serotonin_{1A} receptor binding measured by positron emission tomography with [11C]WAY-100635. Effects of depression and antidepressant treatment. Arch Gen Psychiatry 57: 174-180.
- Sargent P, Williamson DJ, Pearson G, Odontiadis J, Cowen PJ (1997). Effect of paroxetine and nefazodone on 5-HT_{1A} receptor sensitivity. Psychopharmacology 132: 296-302.
- Schlicker E, Göthert M (1998). Interactions between presynaptic alpha₂-autoreceptor and presynaptic inhibitory heteroreceptors on noradrenergic neurons. Brain Res Bull 47: 129-132.
- Serres F, Azorin JM, Valli M, Jeanningros R (1999). Evidence for an increase in functional platelet 5-HT_{2A} receptors in depressed patients using the new ligand [125DOI]. Eur Psychiatry 14: 451-457.
- Sibug RM, Compaan JC, Meijer OC, Van der Gugten J, Olivier B, De Kloet ER (1998). Flesinoxan treatment reduces 5-HT_{1A} receptor mRNA in the dentate gyrus independently of high plasma corticosterone levels. Eur J Pharmacol 353: 207-214.
- Smith CB, García-Sevilla JA, Hollingsworth PJ (1981). α_{2A} -Adrenoceptors in rat brain are decreased after long-term tricyclic antidepressant drug treatment. Brain Res 210: 413-418.
- Stahl S (1994). 5HT_{1A} receptors and pharmacotherapy. Is serotonin receptor down-regulation linked to the mechanism of action of antidepressant drugs? Psychopharmacol Bull 30: 39-43.
- Starke K (1981). Presynaptic receptors. Annu Rev Pharmacol Toxicol 21: 7-30.
- Stockmeier CA, Shapiro LA, Dilley GE, Kolli TN, Friedman L, Rajkowska G (1998). Increase in serotonin-1A autoreceptors in the midbrain of suicide victims with major depression—postmortem evidence for decreased serotonin activity. J Neurosci 18: 7394-7401.
- Subhash MN, Jagadeesh S (1997). Imipramine-induced changes in 5-HT2 receptor sites and inositoltriphosphate levels in rat brain. Neurochem Res 22: 1095-1099.
- Tamotsu S, Schomerus C, Stehle JH, Rosemboom PH, Korf HW (1995). Norepinephrine-induced phosphorylation of the transcription factor CREB in isolated rat pinealocytes: an immunocytochemical study. Cell Tissue Res 282: 219-226.
- Vazquez DM, Eskandari R, Zimmer CA, Levine S, Lopez JF (2002). Brain 5-HT receptor system in the stressed infant rat: implications for vulnerability to substance abuse. Psychoneuroendocrinology 27: 245-272.
- Ventayol P, Busquets X, García-Sevilla JA (1997). Modulation of immunoreactive protein kinase C- α and β isoforms and G proteins by acute and chronic treatments with morphine and other opiate drugs in rat brain. Naunyn-Schmiedeberg's Arch Pharmacol 335: 491-500.
- Yamamoto M, Komori T, Matsumoto T, Zhang K, Miyahara S, Shizuya K et al (2003). Effects of single and repeated prolonged stress on mu-opioid receptor mRNA expression in rat gross hypothalamic and midbrain homogenates. Brain Res 980:
- Young S, Parker PJ, Ullrich A, Stabel S (1987). Down-regulation of protein kinase C is due to an increased rate of degradation. Biochem J 244: 775-779.
- Zhu QS, Chen K, Shih JC (1995). Characterization of the human 5-HT_{2A} receptor gene promoter. J Neurosci 15: 4885-4895.
- Zubieta JK, Ketter TA, Bueller JA, Xu Y, Kilbourn MR, Young EA et al (2003). Regulation of human affective responses by anterior cingulate and limbic mu-opioid neurotransmission. Arch Gen Psychiatry 60: 1145-1153.